

1,2-Diethyl-3-hydroxy-6-phenylthiopyridin-4(1H)-one

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Key indicators

Single-crystal X-ray study

$T = 293\text{ K}$

Mean $\sigma(\text{C}-\text{C}) = 0.004\text{ \AA}$

Disorder in main residue

R factor = 0.047

wR factor = 0.127

Data-to-parameter ratio = 16.4

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

The title compound, $\text{C}_{15}\text{H}_{17}\text{NO}_2\text{S}$, synthesized as an inhibitor for 5-lipoxygenase, comprises the neutral 1,2-diethyl-3-hydroxy-6-phenylthiopyridin-4(1H)-one molecule. The H atom of the hydroxy group and the carbonyl O atom form intermolecular hydrogen bonds with another molecule in a head-to-head fashion. The resulting dimers pack along the b axis and form hydrophobic channels.

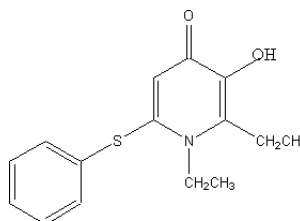
Received 14 January 2002

Accepted 17 January 2002

Online 25 January 2002

Comment

3-Hydroxypyridin-4(1H)-one derivatives demonstrate various pharmacological effects, such as antineoplastic, anti-hypertensive, cardiotoxic, anti-inflammatory and analgesic effects (Hwang *et al.*, 1980; Hershko *et al.*, 1992; Feng *et al.*, 1993; Williams, 1976), and favourable effects in Parkinson's and Thalassemia diseases (Waldmerir *et al.*, 1993; Tondury *et al.*, 1990). In particular, 1,2-dimethyl- and 1,2-diethyl-3-hydroxypyridin-4(1H)-one derivatives have been shown to have a potent chelating effect (Hider *et al.*, 1990). They can effectively remove iron from iron-overloaded animals, including man (Porter *et al.*, 1990, 1994), and have been used clinically. These results prompted us to prepare new 3-hydroxypyridin-4(1H)-one derivatives and assay them for new medicinal effects.



(I)

We have synthesized 6-substituted-*N*-alkyl-3-hydroxypyridin-4(1H)-ones, including the title compound, (I), and examined them for inhibition of 5-lipoxygenase. 5-Lipoxygenase is a cytosolic enzyme which contains a non-heme Fe atom at the active site (Percival, 1991) and catalyses the oxidation of arachidonic acid to leukotrienes, which can cause asthma, inflammatory and rheumatoid arthritis *etc.* We have found that the title compound exhibits strong inhibitory activity on 5-lipoxygenase. Interestingly, the substitution at the 6-position is essential for inhibition of the enzyme. Removal of the substituent at the 6-position, such as in the derivative 1,2-diethyl-3-hydroxypyridin-4(1H)-one, diminished the enzyme inhibitory activity. This is related to both the ability of coor-

dination and hydrophobic discrimination of the title compound.

The carbonyl C—O bond length is 1.259 (2) Å, which is longer than the pyridinone average C=O double bond (1.20 Å) and slightly shorter than those found in 1,2-dimethyl-3-hydroxypyridin-4(1*H*)-one and 1-ethyl-2-methyl-3-hydroxypyridin-4(1*H*)-one (Xiao *et al.*, 1992; Clarke *et al.*, 1992). The phenolic hydroxy C—O distance is 1.354 (2) Å, which is quite close to 1.356 (1) Å, and the C2—C3—C4 bond angle is 114.9 (2)°, which is also close to the value of 114.7 (1)° observed by Xiao *et al.* (1992). The S—C bond lengths are 1.775 (2) and 1.779 (2) Å, and the C—S—C bond angle is 102.52 (9)°. The H atom of the hydroxyl group and the carbonyl O atom of compound (I) are hydrogen bonded to the carbonyl O and the hydroxyl H atom, respectively, from another molecule, resulting in head-to-head dimers. The packing of the dimers generates hydrophobic channels along the *b* axis. Compared with other structurally characterized pyridinone compounds (Nelson *et al.*, 1988), the hydrophobic channels observed in compound (I) possibly result from the addition of the auxiliary phenylthio group.

Experimental

1,2-Diethyl-3-hydroxypyridin-4(1*H*)-one (0.84 g, 5.0 mmol) and Ag₂O (6.0 mmol) were stirred in ethanol (40 ml) at 318 K for 4 h. The solid phase was removed by filtration and the solvent was evaporated. The crude product and thiophenol (0.66 g, 6.0 mmol) were stirred in acetone (20 ml) for 48 h at 308 K. The resulting precipitate was collected by evaporation of the solvent, and was recrystallized from acetone/petroleum ether to afford 0.66 g (2.4 mmol, 48%) of the title compound. Then the compound was dissolved in acetone, and the resulting solution was allowed to stand at room temperature. After 3 d, colorless block crystals of (I) were collected.

Crystal data

C ₁₅ H ₁₇ NO ₂ S	$D_x = 1.247 \text{ Mg m}^{-3}$
$M_r = 275.36$	Mo $K\alpha$ radiation
Monoclinic, $P2_1/n$	Cell parameters from 820 reflections
$a = 12.980 (7) \text{ \AA}$	$\theta = 3.0\text{--}26.6^\circ$
$b = 8.482 (5) \text{ \AA}$	$\mu = 0.22 \text{ mm}^{-1}$
$c = 14.189 (8) \text{ \AA}$	$T = 293 (2) \text{ K}$
$\beta = 110.165 (9)^\circ$	Block, colorless
$V = 1466.4 (14) \text{ \AA}^3$	$0.46 \times 0.26 \times 0.20 \text{ mm}$
$Z = 4$	

Data collection

Bruker CCD area-detector diffractometer	3200 independent reflections
φ and ω scans	2277 reflections with $I > 2\sigma(I)$
Absorption correction: multi-scan (Blessing, 1995)	$R_{\text{int}} = 0.022$
$T_{\text{min}} = 0.906$, $T_{\text{max}} = 0.958$	$\theta_{\text{max}} = 27.2^\circ$
8374 measured reflections	$h = -16 \rightarrow 16$
	$k = -10 \rightarrow 6$
	$l = -18 \rightarrow 17$

Refinement

Refinement on F^2	$w = 1/[\sigma^2(F_o^2) + (0.0454P)^2 + 0.6466P]$
$R[F^2 > 2\sigma(F^2)] = 0.047$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.127$	$(\Delta/\sigma)_{\text{max}} < 0.001$
$S = 1.06$	$\Delta\rho_{\text{max}} = 0.27 \text{ e \AA}^{-3}$
3200 reflections	$\Delta\rho_{\text{min}} = -0.31 \text{ e \AA}^{-3}$
195 parameters	Extinction correction: <i>SHELXL97</i>
H atoms treated by a mixture of independent and constrained refinement	Extinction coefficient: 0.016 (2)

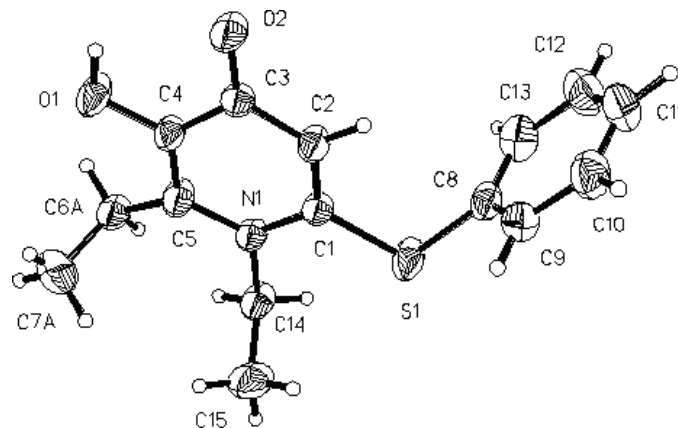


Figure 1

The structure of the title compound, showing 30% probability displacement ellipsoids. The occupancies of the disordered ethyl atoms C6A and C7A is 0.70; the alternative positions (C6B and C7B) have been omitted for clarity.

Table 1

Selected geometric parameters (Å, °).

S1—C8	1.775 (2)	O1—C4	1.354 (2)
S1—C1	1.779 (2)	O2—C3	1.259 (2)
C8—S1—C1	102.52 (9)	O1—C4—C3	118.91 (16)
C2—C3—C4	114.94 (16)	C5—C4—C3	122.39 (18)
O1—C4—C5	118.68 (18)		

Table 2

Hydrogen-bonding geometry (Å, °).

$D\text{—}H\cdots A$	$D\text{—}H$	$H\cdots A$	$D\cdots A$	$D\text{—}H\cdots A$
O1—H1A \cdots O2 ⁱ	0.84 (3)	1.89 (3)	2.638 (2)	147 (3)
O1—H1A \cdots O2	0.84 (3)	2.31 (3)	2.727 (2)	111 (2)

Symmetry code: (i) $2 - x, -y, 1 - z$.

All H atoms bonded to C atoms were placed in idealized positions (C—H = 0.97, 0.96 and 0.93 Å for CH₂, CH₃ and CH, respectively) and refined as riding atoms. The hydroxyl H atom was located from a difference map and was refined. One of the ethyl groups (C6 and C7) is disordered and was refined in two positions C6A/C7A and C6B/C7B.

Data collection: *SMART* (Bruker, 1998); cell refinement: *SMART*; data reduction: *SAINT-Plus* (Bruker, 1999); program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *SHELXTL* (Bruker, 1998); software used to prepare material for publication: *SHELXTL*.

This project is supported by Guangdong Provincial Natural Science Foundation of China (No. 2KM04103S).

References

- Blessing, R. (1995). *Acta Cryst.* **A51**, 33–38.
 Bruker (1998). *SMART* (Version 5.0) and *SHELXTL* (Version 5.1) Bruker AXS Inc., Madison, Wisconsin, USA.

- Bruker (1999). *SAINT-Plus*. Version 6.0. Bruker AXS Inc., Madison, Wisconsin, USA.
- Clarke, E. T., Martell, A. E. & Reibenspies, J. (1992). *Inorg. Chim. Acta*, **196**, 177–183.
- Feng, M. H., Vander, D. L. & Banties, A. (1993). *J. Med. Chem.* **36**, 2822–2827.
- Hershko, C., Godeuk, U. R. & Thuma, P. E. (1992). *J. Inorg. Biochem.* **47**, 267–277.
- Hider, R. C., Taylor, P. D. & Walkinshaw, M. (1990). *J. Chem. Res.* pp. 316–317.
- Hwang, D. R., Proctor, G. R. & Driscoll, S. D. (1980). *J. Pharm. Sci.* **69**, 1074–7076.
- Nelson, W. O., Karpishin, T. B., Rettig, S. J. & Orvig, C. (1988). *Can. J. Chem.* **66**, 123–131.
- Percival, M. D. (1991). *J. Biol. Chem.* **266**, 10058–10062.
- Porter, J. B., Morgan, J. & Hoyes, K. P. (1990). *Blood*, **76**, 2389–2396.
- Porter, J. B., Singh, S. & Hoyes, K. P. (1994). *Adv. Exp. Med. Biol.* **356**, 361–370.
- Sheldrick, G. M. (1997). *SHELXS97* and *SHELXL97*. University of Göttingen, Germany.
- Tondury, P., Kontoghiorghes, G. J. & Ridolfi, A. (1990). *Br. J. Haematol.* **76**, 550–558.
- Waldmerir, P. C., Buchle, A. M. & Steulet, A. F. (1993). *Biochem. Pharm.* **45**, 2417–2421.
- Williams, W. R. H. (1976). *Can. J. Chem.* **54**, 3377–3382.
- Xiao, G. Y., van der Helm, D., Hider, R. C. & Dobbin, P. S. (1992). *J. Chem. Soc. Dalton. Trans.* pp. 3265–3271.